SPECIAL COMMUNICATION

Common Data Elements for Traumatic Brain Injury: Recommendations From the Biospecimens and Biomarkers Working Group

Geoffrey T. Manley, PhD, Ramon Díaz-Arrastia, MD, PhD, Mary Brophy, MD, MPH, Doorje Engel, MD, PhD, Clay Goodman, MD, Katrina Gwinn, MD, Timothy D. Veenstra, PhD, Geoffrey Ling, MD, PhD, Andrew K. Ottens, PhD, Frank Tortella, PhD, Ronald L. Hayes, PhD


Recent advances in genomics, proteomics, and biotechnology have provided unprecedented opportunities for translational research and personalized medicine. Human biospecimens and biofluids represent an important resource from which molecular data can be generated to detect and classify injury and to identify molecular mechanisms and therapeutic targets. To date, there has been considerable variability in biospecimen and biofluid collection, storage, and processing in traumatic brain injury (TBI) studies. To realize the full potential of this important resource, standardization and adoption of best practice guidelines are required to ensure the quality and consistency of these specimens. The aim of the Biospecimens and Biomarkers Working Group was to provide recommendations for core data elements for TBI research and develop best practice guidelines to standardize the quality and accessibility of these specimens. Consensus recommendations were developed through interactions with focus groups and input from stakeholders participating in the interagency workshop on Standardization of Data Collection in TBI and Psychological Health held in Washington, DC, in March 2009. With the adoption of these standards and best practices, future investigators will be able to obtain data across multiple studies with reduced costs and effort and accelerate the progress of genomic, proteomic, and metabolomic research in TBI.

Key Words: Biological Markers; Brain Injuries; Rehabilitation. © 2010 by the American Congress of Rehabilitation Medicine

Because genomic and proteomic research in TBI is still very much an emerging field, our goal was not to recommend assays of specific protein biomarkers or polymorphisms as core data elements. The aim of the working group was to provide recommendations for core biospecimen and biomarker data elements for TBI research and develop best practice guidelines to standardize the quality and accessibility of these specimens. We were also charged with establishing consensus for the core specimen data elements that should be collected for every TBI study and to identify supplemental and emerging data elements for more advanced and extended studies.

Approach

Multiple group conference calls were held as well as a number of smaller group calls and e-mail interactions. There was universal agreement on the need for standardization and development of best practices for collecting, processing, and storing biospecimens and biofluids. Past, current, and future efforts in civilian and military TBI biomarker genomics and proteomics research were reviewed and discussed.

Two series of observations support the fundamental hypothesis that genetic factors significantly influence functional outcome after TBI. The first of these is the finding that inheritance of the APOE-e4 allele is associated with poor outcome after TBI.1 The effect size of inheritance of one APOE-e4 allele is modest, with the odds ratio of an unfavorable outcome ranging from 3.57 to 13.93. While these and other studies have primarily focused on 6-month outcome,3 there is evidence that the APOE-e4 genotype may also be associated with long-term outcome and cognitive decline after TBI.4 Thus, this polymorphism may be relevant in the patient with chronic as well as acute TBI. Polymorphisms in the interleukin-1 system have not so far been replicated.5 It is likely that the small sample sizes used in studies to date are a factor in false-negative results but the need for large sample sizes to address this issue continues.6 Currently, the most effective treatments for TBI are focused on acute management and the use of antioxidants and anti-inflammatory agents to reduce secondary injury.7 However, there is limited evidence regarding the effectiveness of these agents.8 New treatments are needed to improve functional outcomes and quality of life for patients with TBI.9,10

Technological advances in genomics, proteomics, and biofluid analysis provide unprecedented opportunities to detect and classify injury and to identify molecular mechanisms and therapeutic targets. To date, there has been considerable variability in biospecimen and biofluid collection, storage, and processing in TBI studies. To realize the full potential of this important resource, standardization and adoption of best practice guidelines are required to ensure the quality and consistency of these specimens. The aim of the Biospecimens and Biomarkers Working Group was to provide recommendations for core data elements for TBI research and develop best practice guidelines to standardize the quality and accessibility of these specimens. Consensus recommendations were developed through interactions with focus groups and input from stakeholders participating in the interagency workshop on Standardization of Data Collection in TBI and Psychological Health held in Washington, DC, in March 2009. With the adoption of these standards and best practices, future investigators will be able to obtain data across multiple studies with reduced costs and effort and accelerate the progress of genomic, proteomic, and metabolomic research in TBI.

List of Abbreviations

APOE-e4 apolipoprotein E4
CDE common data element
COMT catechol O-methyltransferase
dna deoxyribonucleic acid
EDTA ethylenediaminetetraacetic acid
EVD external ventricular drain
LP lumbar puncture
PBMC peripheral blood mononuclear cell
RBC red blood cell
TBI traumatic brain injury

From the University of California San Francisco, San Francisco, CA (Manley); University of Texas Southwestern Medical School, Dallas, TX (Díaz-Arrastia); Department of Veterans Affairs Cooperative Studies Program, Boston, MA (Brophy); Heidelberg University Hospital, Heidelberg, Germany (Engel); Baylor College of Medicine, Houston, TX (Goodman, Gwinn); Science Applications International Corporation-Frederick Inc, National Cancer Institute at Frederick, Frederick, MD (Veenstra); Uniformed Services University, Bethesda, MD (Ling); Walter Reed Army Institute of Research, Silver Spring, MD (Tortella); Departments of Anatomy and Neurobiology, and Biochemistry, Virginia Commonwealth University, Richmond, VA (Ottens); Banyan Biomarkers Inc, Alachua, FL (Hayes).

A commercial party having a direct financial interest in the results of the research supporting this article has conferred or will confer a financial benefit on the author or one or more of the authors. R. L. Hayes is founder and president of Banyan Biomarkers Inc. The views, opinions, or assertions contained herein are the private views of the authors and do not necessarily reflect those of the agencies or institutions with which they are affiliated, including the U.S. Department of Veterans Affairs, U.S. Department of Defense, Department of the Army, the U.S. Department of Health and Human Services, the National Institutes of Health, the National Institute of Mental Health, and the Uniformed Services University of the Health Sciences. This work is not an official document, guidance, or policy of the U.S. Government, nor should any official endorsement be inferred.

Correspondence to Geoffrey T. Manley, MD, PhD, University of California, San Francisco, Department of Neurosurgery, 1001 Potrero Ave, Building 1, Room 101, San Francisco, CA 94110, e-mail: manleyg@neurosurg.ucsf.edu. Reprints are not available from the author.

0003-9993/10/9111-00370$36.00/0 doi:10.1016/j.apmr.2010.05.018

Arch Phys Med Rehabil Vol 91, November 2010
negative and even false-positive findings. Although APOE-e4 appears to be the most likely candidate for the first “Core” genomic biomarker for TBI, future studies with sufficient patient numbers and comprehensive phenotyping are needed. There are also recent examples of discovery-driven proteomic studies focused on finding effective biomarkers for TBI. Mass spectrometry has been the primary proteomic technology and has revolutionized the ways that biomarker discovery is being conducted in this field.

Genomic biomarkers may also be relevant to psychologic health issues associated with TBI. For example, the COMT gene plays a key role in the degradation of dopamine in the frontal cortex. The Val158Met polymorphism in the COMT gene has been associated with neuropsychiatric phenotypes and cognition. The Val158Met genotype has also been associated with executive functioning in patients with TBI. Recent studies have also identified an association between serotonin transporter genotype and posttraumatic stress disorder, suggesting that this and other polymorphisms may help us understand the complicated interrelationship between TBI and psychologic health.

It was the consensus of the group that while there is significant potential for large-scale genomics and proteomics in TBI, there is sufficient variability in TBI biospecimen collection, processing, and storage that must be addressed to advance the field. Prospective clinical trials are an ideal setting in which to collect DNA and biological fluids in sufficiently large sample sizes associated with carefully collected clinical data to allow successful genomic and proteomic studies. A due diligence process was carried out to explore similar efforts in other fields. A number of groups have already developed best practice guidelines for biospecimen resources. These documents provide general principles that can be used to guide the design of studies in which biospecimens will be analyzed. There was general agreement that much of the groundwork for the TBI biospecimen working group had been provided by these prior efforts. It was acknowledged that the group should not reinvent the wheel, but it was also recognized that there are issues particular to TBI that should be considered and addressed. The group felt that a document that built on past efforts and incorporated issues relevant to TBI would be an important contribution to the literature and the research community. Based on the members' individual expertise, the participants divided into subgroups to address various aspects of biofluid and biomarker specimen collection, storage, and processing. Preliminary recommendations were developed through e-mail interactions with the focus groups and discussed in person with the working group members at the interagency workshop on Standardization of Data Collection in TBI and Psychological Health held in Washington, DC, in March 2009. The refined draft recommendations and best practices were presented to stakeholders and other working groups during the workshop for additional input. These comments and suggestions have also been incorporated in the current recommendations. The product of the Biospecimens and Biomarkers Working Group is, by its nature, different from the other TBI CDE Working Group articles in that the focus is best practices with step-by-step protocols to standardize sample collection, processing, and storage.

**RECOMMENDATIONS FOR COMMON DATA ELEMENTS FOR BIOSPECIMENS AND BIOMARKERS**

The following are the group’s recommendations for core specimen data elements that should be collected for all studies. The collection of supplemental and emerging data elements recommended for more advanced and extended studies should be encouraged, particularly in studies that are directed at a molecular mechanism that the biomarker measures.

1. Core data element recommendations
   a. Collection of DNA sample for genomic analysis
   b. Collection of acute (<24h) plasma sample for proteomic and metabolomic analyses

2. Supplemental data element recommendations
   a. Collection of serial plasma and serum samples for proteomic analysis
   b. Collection of cerebrospinal fluid samples for proteomic analysis

3. Emerging data element recommendations
   a. Collection of cerebral microdialysis samples
   b. Collection of PBMCs for gene and protein expression studies

The TBI CDE Working Group recommendations for DNA guidelines for genomic analyses are presented in appendix 1. The timing of DNA collection is less critical, even in patients who have received a blood transfusion. Recommendations for plasma and serum guidelines for proteomic and metabolomic analyses are shown in appendix 2. The timing for acquiring these samples is more complicated. An acute plasma sample (defined as <24h) was recommended by the Working Group as “Core” for all TBI studies to provide the opportunity for the identification of diagnostic and predictive biomarkers in large series of patients. However, the frequency and duration of serial sample collection is biomarker-dependent and cannot be standardized at this time. Information regarding cerebrospinal fluid guidelines and microdialysis guidelines can be found in appendices 3 and 4, respectively. Each of these best practice guidelines addresses the acquisition, processing, and storage of the samples in sufficient detail to promote standardization.

**FUTURE DIRECTIONS**

Although these recommendations and best practices are voluntary, their adoption will lead to a new standardization that will advance TBI research. To demonstrate the utility of these recommendations and best practices, we propose a pilot study to examine the feasibility of multicenter biomarker CDE collection across the broad spectrum of TBI. Implementation of biospecimen and biomarker best practices will also require outreach and education to inform and engage the TBI research community and solicit comments and feedback. The working group believes that the recommendations and best practices for TBI biospecimens and biomarkers will continue to evolve as the field advances and potential biomarkers will one day become core CDEs. Thus, there will be a need for ongoing support for the TBI CDE effort.

**APPENDIX 1: DNA GUIDELINES FOR GENOMIC ANALYSES**

I. Acquisition of blood biospecimens
   A. Blood is collected through venipuncture by appropriately trained personnel.
   B. For isolation of genomic DNA from whole blood, collection of 5 to 20mL using EDTA-containing Vacutainer tubes is suitable. Use of citrate or heparinized tubes is also suitable.
   C. Invert the sample 8 to 10 times to ensure proper mixture of blood and anticoagulant.
   D. For isolation of PBMCs for future preparation of lymphoblastoid cell lines or gene expression studies, col-
APPENDIX 1 (Cont’d): DNA GUIDELINES FOR GENOMIC ANALYSES

III. Local storage

A. Whole blood (in EDTA or LeucoPREP tubes) should be maintained at room temperature until transfer to the laboratory for processing. Studies have shown that packed cell volume starts decreasing with EDTA addition as early as 1 hour postcollection, so it is important to process the specimen in a time-efficient manner.

B. Transport the original, unfrozen Vacutainer without breaking the seal to the designated local genomics laboratory (if available). The Vacutainer system best preserves the integrity of the blood sample if it is not broken.

C. Because the expense of overnight shipping is justifiable only when highly specialized procedures such as preparation of lymphoblastoid cell lines are planned, the most cost-effective approach when such procedures are not required is to aliquot and freeze whole blood locally for DNA isolation later.
   i. Storage in aliquots of 1-mL to 2-mL freezer-safe containers (ie, Cryovials) is most convenient.
   ii. Multiple aliquots should be prepared in case of mishaps during shipment or DNA isolation. This also minimizes the freeze-thaw cycles should there be a need for multiple analyses at interval times.

D. DNA isolation from whole blood. It is most cost-efficient to isolate DNA from multiple (20–100 samples) at a time. This is most efficiently done by having each of the clinical sites store samples locally until a suitable number have been collected (ie, 10–20 samples). These can then be shipped in a single overnight shipment to a central laboratory experienced in collection and banking of samples.

E. Preparation of lymphoblastoid cell lines. Overnight shipment of LeucoPREP tubes to a laboratory experienced in the viral transformation and preparation of lymphoblastoid lines.

III. Local storage

A. Appropriate and complete documentation surrounding biospecimen collection, processing, and storage are essential and will influence the quality of research data to be obtained.

B. Samples should be placed in nonfrost-free freezers at or below −80°C. Frost-free freezers go through freeze-thaw cycles that further damage the specimen.

C. Avoid any thawing of frozen samples. Thawing of frozen samples of whole blood results in release of deoxyribonucleases that destroy the DNA.

D. Centers at which samples are stored should institute a back-up plan for freezer failure (eg, alternate power source, dry ice, or liquid nitrogen). An appropriate alarm system to support freezers for longtime storage is essential.

E. Lymphoblastoid cell lines must be stored in liquid nitrogen. The Coriell Institute provides storage for samples collected in National Institutes of Health-sponsored studies (http://www.coriell.org).

F. An inventory system should be established for tracking provenance of samples, including the time of collection, processing, storage, and quality control procedures carried out on each sample.

IV. Shipping

A. Deoxyribonucleases degradation begins after 2 to 3 days at room temperature, so fresh blood samples should be shipped to the processing site within hours if possible. If the genomics laboratory is not within reach of local transport, frozen blood samples may be shipped through a designated agency.

B. Dry ice or cold packs must accompany the frozen specimen during air shipment. When dry ice is used, the transport time should be minimized given that dry ice sublimates at a rate of 2 to 5 kg per 24 hours depending on the insulation of the shipment container.

C. Consult the local agency for proper shipping options and certified transport materials. The International Air Transportation Association (http://www.iata.org) and the U.S. Department of Transportation (http://www.dot.gov) have legal requirements governing the packaging, labeling, and shipping of biospecimens.
   i. Category A infectious substances are capable of causing permanent disability or life-threatening or fatal disease in humans or animals when exposure occurs.
   ii. Category B infectious substances (also “diagnostic specimens” or “clinical specimens”) are infectious but do not meet the standard for category A inclusion.
   iii. Exempt patient specimens have a minimal likelihood of containing pathogens.

D. Temperature loggers can be used to monitor temperature in shipments of samples to provide confirmation and assurance that samples have been maintained at appropriate temperatures.

V. Central storage

A. Appropriate and complete documentation surrounding biospecimen collection, processing, storage, and shipping from the individual sites is essential and will influence the quality of the multicenter research data to be obtained.

B. Bar code identification of samples with an automated date and time stamp is recommended.

C. A formal plan for sharing the central biospecimen resource is recommended.

D. The Central Storage Bank should maintain information of laboratories where the samples have been sent to avoid duplicative genotyping and inadvertent repetitive reporting of data from the same patient.

E. The Central Storage Bank should also maintain information regarding any stipulations regarding informed consent for the use of the samples. For example, in some studies, participants may provide permission for their samples to be used only for studies on TBI.

APPENDIX 2: PLASMA AND SERUM GUIDELINES FOR PROTEOMIC ANALYSES

I. Acquisition of blood biospecimens

A. For severely injured patients, blood is collected via vascular access catheters that have been placed as part
APPENDIX 2 (Cont’d): PLASMA AND SERUM GUIDELINES FOR PROTEOMIC ANALYSES

of the patients’ routine medical care. For other patients, trained personnel collect blood through venipuncture.

B. For most purposes, 5 to 10mL whole blood will be collected using a Vacutainer system. It should be noted that the use of glass tubes can lead to low values for certain analytes. For the most generalizable purposes, polypropylene Vacutainer and subsequent storage tubes are recommended.

II. Local processing (serum)

A. Blood samples should be collected in vacutainers that contain no anticoagulant for the processing of serum.
B. Samples should be sat upright at room temperature for 30 minutes to allow for clotting. Then they should be spun at 4000rpm at room temperature for 5 to 7 minutes. The cleared serum should be pipetted and stored in aliquots of 1 to 2mL.
C. Record the volume of each aliquot.

III. Local processing (plasma)

A. Blood samples should be collected in vacutainers that contain EDTA when preparing plasma. Previous research suggests that EDTA is the preferred anticoagulant because others may interfere with analyte detection.19
   i. The distinction between K2EDTA and K3EDTA and their concentrations should be assessed. See Goossens et al20 for more details.
B. Transport the original, unfrozen blood sample to the designated local proteomics laboratory as soon as possible. Freezing has significant adverse effects on plasma and its proteomic elements.
C. If specimens must be stored before processing, the samples should sit on ice for 5 to 10 minutes. Samples should be spun at 4000rpm at room temperature for 5 to 7 minutes. Plasma should be pipetted and stored in aliquots of 500 μL to 2mL.
D. Record the volume of each aliquot.

IV. Local documentation and storage

A. Appropriate and complete documentation surrounding biospecimen collection, processing, and storage are essential and will influence the quality of research data to be obtained.
B. Bar code identification of samples with an automated date and time stamp is recommended.
C. Samples should be placed in nonfrost-free freezers at or below –80°C. Frost-free freezers go through freeze-thaw cycles that further damage the specimen.
D. Centers at which samples are stored should institute a back-up plan for freezer failure (eg, dry ice or liquid nitrogen). An appropriate alarm system to support freezers is essential and will influence the quality of research data to be obtained.
E. An inventory system should be established for tracking provenance of samples, including the time of collection, processing, storage, and quality control procedures carried out on each sample.

V. Shipping

A. Dry ice or cold packs must accompany the frozen specimen during air shipment. When dry ice is used, the transport time should be minimized given that dry ice sublimes at a rate of 2 to 5 kg per 24 hours depending on the insulation of the shipment container.
B. Consult the local agency for proper shipping options and certified transport materials. The International Air Transportation Association (http://www.iata.org) and the U.S. Department of Transportation (http://www.dot.gov) have legal requirements governing the packaging, labeling, and shipping of biospecimens.
   i. Category A infectious substances are capable of causing permanent disability or life-threatening or fatal disease in humans or animals when exposure occurs.
   ii. Category B infectious substances (also “diagnostic specimens” or “clinical specimens”) are infectious but do not meet the standard for Category A inclusion.
   iii. Exempt patient specimens have a minimal likelihood of containing pathogens.
D. Temperature loggers can be used to monitor temperature in shipments of samples to provide confirmation and assurance that samples have been maintained at appropriate temperatures.

VI. Central storage (see appendix 1)

APPENDIX 3: CSF GUIDELINE

I. Acquisition of CSF from an EVD

A. Document whether continuous or intermittent (catheter opened only in response to intracranial hypertension) fluid drainage is administered. Drainage method has been shown to alter CSF protein concentration.21
B. Draw CSF directly from ventriculostomy catheter.
C. Target CSF collection within the first 24 hours of admission, recording time from TBI and time of day. Ideally, the first collection should be as close to the TBI as feasible (eg, 6h post-TBI), at a minimum frequency of every 6 hours for sufficient biokinetic studies.
D. Collect 5mL of CSF in 1-mL fractions and place in ice bath. The first 1 or 2 fractions are sent for clinical laboratory analysis for cell count and protein and glucose measurements. Blood contamination of CSF is a significant confounder. Protein concentrations are 400-fold greater in plasma than CSF.22 CSF is considered blood-contaminated if RBC counts are greater than 10 cells/μL or if hemoglobin levels are greater than 30pg/mL CSF.23 Brain specific proteins are typically present at low concentrations in CSF, with 80% of normal CSF protein mass originating from plasma.24
E. Appropriate CSF control samples may be available from hydrocephalic patients who undergo ventriculoperitoneal shunt placement and had CSF collected intraoperatively, or patients with unruptured subarachnoid hemorrhage who had CSF drawn intraoperatively.25

II. Acquisition of CSF from an LP

A. In patients who are unlikely to receive a ventriculostomy, CSF can be accessed by a less invasive LP. A greater number of LP collected control samples are available; however, comparison with EVD CSF is discouraged given a 2.5-fold lower protein concentration than in LP CSF.26
APPENDIX 3 (Cont’d): CSF GUIDELINE

B. Atraumatic spinal needle LP kits should be used to minimize risk of post-LP headache. Draw with a sterile polypropylene syringe or allow flow under gravity.

C. Target CSF collection within the first 24 hours after admission, recording time from TBI and time of day. Ideally, the first collection should be as close to the TBI as feasible (eg. 6h post-TBI) at a minimum frequency of every 6 hours for sufficient biokinetic studies.

D. Collect 1-mL fractions and place in ice bath, with a maximum of 25mL a time point. Send the first 2mL for clinical laboratory analysis. It is important to match fractions when comparing across patients because protein concentration varies depending on the draw volume.27

E. Additives or preservatives may be combined with CSF – 80°C to minimize proteolytic breakdown.29 Avoid repeated freeze and thawing cycles and storage at –20°C.30

III. Processing and storage of CSF

A. Transport CSF on ice and process immediately after collection because significant cell lysis contamination will occur within 1 hour.

B. Collect CSF samples into low protein binding polypropylene tubes (eg, Eppendorf brand LoBind tubes†). Avoid polystyrene and glass tubes, which will result in significant protein loss.28

C. Centrifuge CSF at 2000 rpm for 24 to 48 hours to minimize risk of headache.

D. Patient should rest in a recumbent position for 1 hour post-LP, receive liberal fluid intake, and avoid exertion for 24 to 48 hours to minimize risk of headache.

APPENDIX 4: CEREBRAL MICRODIALYSIS GUIDELINES

III. Microdialysate characteristics

A. Either of the following microdialysate fluids is acceptable:
   i. Artificial CSF
   ii. Sterile medical-grade normal saline

B. The composition and source of the microdialysate must be specified in all data reporting.

IV. Sample acquisition

A. The microdialysate flow rate should be 0.3 µL/min so that it is theoretically possible to collect 18 µL during 1 hour. This flow rate assures near 100% analyte recovery for common low-molecular-weight analytes.

B. Because of the extremely small sample volumes, the samples are sensitive to evaporation even though they are sealed in the microvials, and because microvials may contain different volumes, the effect of evaporation can vary between the samples. Characteristics of the collecting containers must be reported.

C. Samples should be analyzed and/or stored promptly.

V. Analytes

A. The minimal essential data set consists of glucose, pyruvate, and lactate, with use of the calculated lactate/pyruvate ratio.

B. Measurement of glycerol and glutamate is recommended.

C. Determination of other analytes including markers of neuroinflammation such as cytokines, nitrate and nitrite as surrogates for nitric oxide, structural proteins such as glial fibrillary acidic protein, neurofilament, and tau and stress reactants such as beta-amyloid are all experimental. Analysis of these and other potential biomarkers should be encouraged but not required.

VI. Analyte stability during storage

A. The essential low-molecular-weight analytes are stable at –70°C. The samples should be stored at this temperature in containers designed to minimize evaporative losses.

B. Analytes, particularly pyruvate, may not be stable at –20°C. Storage at this temperature is not acceptable with the exception of temporary holding for not more than 3 days.

C. Determination of other analytes including markers of neuroinflammation such as cytokines, nitrate and nitrite as surrogates for nitric oxide, structural proteins such as GFAP, neurofilament, and tau and stress reactants such as beta-amyloid are all experimental. Analysis of these and other potential biomarkers should be encouraged but not required.

VII. Handling of samples at the time of analysis

A. If the samples have been frozen, then they must be thawed prior to analysis. It is important to recognize physicochemical events that may occur in the process of thawing that might cause analytic errors. As the samples thaw, the liquid phase will initially contain a very high concentration of salt and analytes. As the thawing progresses, the solution will be diluted by the melting ice. During this process, there is a risk that the thawed samples are nonhomogeneous; therefore, it is recommended that the samples be thawed and then agitated or centrifuged to assure homogeneous distribution of analyte.
APPENDIX 4 (Cont’d): CEREBRAL MICRODIALYSIS GUIDELINES

B. It may be desirable to thaw the samples rapidly in a heating
cupboard at +40°C for about 10 minutes. Longer times
and/or higher temperatures should not be used because
these may result in a risk for unacceptable evaporation.

C. Stored samples may be assayed using the batch analysis
systems. However, if the low-volume samples sit for too
long in the analyzer prior to analysis, unacceptable evap-
oration may occur. Calibration samples should be inter-
spersed in the batch to detect a systematic elevation in
analyte levels caused by evaporative loss.

VIII. Microdialysis data reporting

A. Analyte concentrations should be reported in Interna-
tional System (SI) units.

B. Ratios such as the lactate/pyruvate ratio are devoid of units.

References

1. Teasdale GM, Nicoll JAR, Murray G, Fiddes M. Association of
apolipoprotein E polymorphism with outcome after head injury.

genotype predicts a poor outcome in survivors of traumatic brain

analysis of APOE4 allele and outcome after traumatic brain in-

4. Isoniemi H, Tonovuo O, Portin R, Himalan M, Kairisto V. Out-
come of traumatic brain injury after three decades—relationship to

5. Hadjigeorgiou GM, Paterakis K, Dardiotis E, et al. IL-1RN and
IL-1B gene polymorphisms and cerebral hemorrhagic events after

6. Diaz-Arrastia R, Baxter VK. Genetic factors in outcome after
traumatic brain injury: what the Human Genome Project can teach

7. Prieto DA, Ye X, Veenstra TD. Proteomic analysis of traumatic
2005;8:283-91.

approaches to studying traumatic brain injury. Prog Brain Res
2007;161:401-18.

Svennerholm L. Protein analyses in cerebrospinal fluid. I. Influ-
ence of concentration gradients for proteins on cerebrospinal

10. Teasdale GM, Nicoll JAR, Murray G, Fiddes M. Association of
apolipoprotein E polymorphism with outcome after head injury.

Val158Met genotype with executive functioning following traumatic

12. De Frias CM, Straub RE. Effect of COMT Val108/158Met genotype on
posttraumatic stress disorder diagnosis in 2 independent popula-

for studies of disease-associated protein biomarkers. Mol Cell

for the standardization of cerebrospinal fluid collection and bio-

15. National Cancer Institute best practices guidelines for biospeci-
men resources. Prepared by the National Cancer Institute, Na-
tional Institute of Health, and U.S. Department of Health and

16. International Society for Biological and Environmental Repositories.
Best practices for repositories I: collection, storage, and
retrieval of human biological materials for research. Cell Preserv

17. Organization for Economic Co-Operation and Development.
OECD best practice guidelines for biological resource centres.

human tissue repositories: “best practices” for a biospecimen
resource for the genomic and proteomic era. Santa Monica:
RAND Corp; 2003.

ization of measurement of beta-amylloid (1-42) in cerebrospinal

20. Goossens W, Van Duppen V, Verwilghen RL, K2- or K3-EDTA:
the anticoagulant of choice in routine haematology? Clin Lab

intermittent cerebrospinal fluid drainage after severe traumatic
brain injury in children: effect on biochemical markers. J Neu-

22. Maurer MH. Proteomics of brain extracellular fluid (ECF) and

23. Zhang J. Proteomics of human cerebrospinal fluid—the good,

KE. Peptide mapping of proteins in human body fluids using
electrospray ionization fourier transform ion cyclotron resonance

25. Pineda JA, Lewis SB, Valadka AB, et al. Clinical significance of
all-spectrin breakdown products in cerebrospinal fluid after se-

26. Huhner AF, Biringer RG, Amato H, Fonteth AN, Harrington MG.
Protein analysis in human cerebrospinal fluid: physiological aspects,

Svennerholm L. Protein analyses in cerebrospinal fluid. I. Influ-
ence of concentration gradients for proteins on cerebrospinal


29. Wagner AK, Ren D, Conley YP, et al. Sex and genetic associa-
tions with cerebrospinal fluid dopamine and metabolite production

30. Carrette O, Burkhard PR, Hughes S, Hochstrasser DF, Sanchez
JC. Truncated cystatin C in cerebrospinal fluid: technical [cor-

31. Shores KS, Knapp DR. Assessment approach for evaluating high
abundance protein depletion methods for cerebrospinal fluid (CSF)

Suppliers

a. Becton Dickinson Labware, 1 Becton Dr, Franklin Lakes, NJ 07417.

b. GE Healthcare, 800 Centennial Ave, #1 Piscataway, NJ 08854-3930.

c. Eppendorf AG, Barkhausenweg 1, 22339 Hamburg 22331, Ham-
burg, Germany.